

Associations of Perfluoroalkyl Substances with Incident Natural Menopause: The Study of Women's Health Across the Nation

Ning Ding,¹ Siobán D. Harlow,¹ John F. Randolph Jr,² Antonia M. Calafat,³ Bhramar Mukherjee,⁴ Stuart Batterman,^{5,6} Ellen B. Gold,⁷ and Sung Kyun Park^{1,5}

¹Department of Epidemiology, School of Public Health, University of Michigan, Ann Arbor, Michigan 48109; ²Department of Obstetrics and Gynecology, School of Medicine, University of Michigan, Ann Arbor, Michigan 48109; ³Division of Laboratory Sciences, National Center for Environmental Health, Centers for Disease Control and Prevention, Atlanta, Georgia 30333; ⁴Department of Biostatistics, School of Public Health, University of Michigan, Ann Arbor, Michigan 48109; ⁵Department of Environmental Health Sciences, School of Public Health, University of Michigan, Ann Arbor, Michigan 48109; ⁶Department of Civil and Environmental Engineering, College of Engineering, University of Michigan, Ann Arbor, Michigan 48109; and ⁷Department of Public Health Sciences, University of California, Davis, School of Medicine, Davis, California 95817

ORCID numbers: 0000-0001-5244-2909 (N. Ding); 0000-0001-9981-6250 (S. K. Park).

Context: Previous epidemiologic studies of per- and polyfluoroalkyl substances (PFASs) and menopausal timing conducted in cross-sectional settings were limited by reverse causation because PFAS serum concentrations increase after menopause.

Objectives: To investigate associations between perfluoroalkyl substances and incident natural menopause.

Design and Setting: A prospective cohort of midlife women, the Study of Women's Health Across the Nation, 1999-2017.

Participants: 1120 multiracial/ethnic premenopausal women aged 45-56 years.

Methods: Serum concentrations of perfluoroalkyls were quantified by high-performance liquid chromatography isotope dilution tandem mass spectrometry. Natural menopause was defined as the bleeding episode prior to at least 12 months of amenorrhea not due to surgery or hormone use. Cox proportional hazards models were used to calculate hazard ratios (HRs) and 95% confidence intervals (CIs).

Results: Participants contributed 5466 person-years of follow-up, and 578 had incident natural menopause. Compared with the lowest tertile, women at the highest tertile of baseline serum concentrations had adjusted HR for natural menopause of 1.26 (95% CI: 1.02-1.57) for n-perfluorooctane sulfonic acid (n-PFOS) ($P_{trend} = .03$), 1.27 (95% CI: 1.01-1.59) for branched-PFOS ($P_{trend} = .03$), and 1.31 (95% CI: 1.04-1.65) for n-perfluorooctanoic acid ($P_{trend} = .01$). Women were classified into four clusters based on their overall PFAS concentrations as mixtures: low, low-medium, medium-high, and high. Compared with the low cluster, the high cluster had a HR of 1.63 (95% CI: 1.08-2.45), which is equivalent to 2.0 years earlier median time to natural menopause.

ISSN Print 0021-972X ISSN Online 1945-7197

Printed in USA

© Endocrine Society 2020. All rights reserved. For permissions, please e-mail: journals.permissions@oup.com

Received 3 March 2020. Accepted 20 May 2020.

First Published Online 3 June 2020.

Corrected and Typeset 11 August 2020.

Abbreviations: BMI, body mass index; CI, confidence interval; FMP, final menstrual period; HR, hazard ratio; HT, hormone therapy; IQR, interquartile range; LOD, limit of detection; MPS, Multi-Pollutant Study; PFAS, per- and polyfluoroalkyl substance; PFHxS, perfluorohexane sulfonic acid; PFOA, perfluorooctanoic acid; PFOS, perfluorooctane sulfonic acid; PFNA, perfluorononanoic acid; PPAR, peroxisome proliferator-activated receptor; SWAN, Study of Women's Health Across the Nation.

Conclusion: This study suggests that select PFAS serum concentrations are associated with earlier natural menopause, a risk factor for adverse health outcomes in later life. (*J Clin Endocrinol Metab* 105: e3169–e3182, 2020)

Freeform/Key Words: per- and polyfluoroalkyl substances (PFAS), endocrine-disrupting chemicals, natural menopause, midlife women

Menopause marks the cessation of ovarian function, and its timing has physiologic impacts beyond the reproductive system, affecting the overall health of midlife women (1, 2). Earlier age at the final menstrual period (FMP) has been associated with an increased risk of overall mortality (3–5), cardiovascular disease (6, 7), cardiovascular death (3, 8, 9), low bone mineral density (10) and osteoporosis (11), and other chronic conditions (12). Ovarian aging reflects the combined effects of genetic factors, sociodemographics, and lifestyle and health characteristics (13–15). Although the etiology of premature menopause (before age 40 years) and early menopause (before age 45 years) is not fully understood, accumulating evidence has suggested that certain environmental exposures may play an important role in the acceleration of ovarian aging (16–18).

Per- and polyfluoroalkyl substances (PFASs) are a family of anthropogenic environmentally persistent chemicals, some of which also persist in the human body, that have been widely used in many industrial and consumer products, such as nonstick cookware (19, 20), food packaging (21–23), outdoor apparel (24, 25), and aqueous film-forming foams (26–29). These compounds, especially the most studied perfluorooctanoic acid (PFOA) and perfluorooctane sulfonic acid (PFOS), have been identified as plausible endocrine disruptors with the potential to cause reproductive disturbances (30, 31). The potential for reproductive impact is supported by findings from animal toxicology studies with effects on female reproduction, including altered ovarian function, histopathologic changes in the reproductive tract, and ovarian cell steroidogenesis (32–34), likely through the activation of various transcriptional factors, such as peroxisome proliferator-activated receptors (PPARs) (35, 36). However, extrapolations of findings from animal studies to the potential effects of PFASs on human ovarian health are clearly limited, given the species-specific toxicokinetics, metabolism, and tissue distributions of PFASs (37).

Although 3 human studies have examined the associations of natural menopause with PFOS, PFOA, perfluorononanoic acid (PFNA), and perfluorohexane sulfonic acid (PFHxS), the results have been inconsistent. A cross-sectional study of mid-Ohio Valley residents found that earlier age at natural menopause was associated with higher concentrations of PFOA and PFOS

(38), whereas using data from the National Health and Nutrition Examination Survey (NHANES), Taylor et al. observed a significant relationship of earlier natural menopause with PFHxS but not with PFOA, PFOS, and PFNA (39). These studies also raised concerns about reverse causation, in that it is unclear whether PFAS exposure contributed to earlier menopause, or cessation of PFAS excretion via cessation of menstruation led to increased serum concentrations of PFASs in women (39–42).

A retrospective cohort study reported no association between PFOA exposure and natural menopause (43). That study relied on recalled information on age at menopause that had occurred on average >10 years prior to the interview. It is difficult to ascertain the precise timing of FMP without longitudinal observations of menstrual cycles (44). Potential recall bias may have reduced the accuracy of reported age at natural menopause and presumably biased the study results toward the null (43). Annual interviews can determine relatively accurate estimates of FMP, and a prospective cohort design with a large, diverse population can provide insights regarding causality that can be more generalizable.

We, therefore, examined the associations between perfluoroalkyl substances and incidence of natural menopause in the multiracial/ethnic sample of women who were premenopausal at baseline from a prospective cohort, namely the Study of Women's Health Across the Nation (SWAN). Women were followed every year from 1999–2010 and every other year from 2011–2017. We also assessed whether the relationship differed by racial/ethnic groups and evaluated the combined effects of chemical mixtures on natural menopause.

Materials and Methods

Study design

The SWAN cohort, a multiracial/ethnic, longitudinal study, was designed to characterize physiologic and psychosocial changes that occur during the menopausal transition to observe their effects on subsequent risk factors for age-related chronic diseases, as previously described (45). A total of 3302 premenopausal women aged 42 to 52 years at baseline were recruited from 7 study sites, including Boston, MA; Chicago, IL; Detroit, MI; Los Angeles, CA; Newark, NJ; Oakland, CA; Pittsburgh, PA. Eligible participants had to have an intact uterus, at least 1 menstrual period in the prior 3 months,

and not have taken hormone medications within the prior 3 months. Participants self-identified as non-Hispanic White women or 1 designated minority group, including Black, Chinese, Hispanic, and Japanese in a proportion for each site. Data and specimens were collected every year from 1999 to 2010 and every other year from 2011 to 2017. The institutional review board at each participating site approved the study protocol, and all participants provided written, signed informed consent.

The SWAN Multi-Pollutant Study (MPS) was initiated in 2016, using the SWAN follow-up visit 03 (V03, 1999-2000) as the baseline to examine the potential health effects of multiple environmental chemicals, including PFAS, polychlorinated biphenyls, organochloride pesticides, polybrominated diphenyl ethers, metals, phenols, phthalates, and organophosphate pesticide among midlife women. The study design of the SWAN MPS is shown in the digital repository (all supplementary material and figures are located in a digital research materials repository (46)). We used repository serum and urine samples collected at SWAN V03, considered the MPS baseline for environmental exposure assessments. Of 2694 women enrolled at SWAN V03, we did not include women from Chicago ($n = 368$) and Newark ($n = 278$), because urine samples were not available at these 2 sites. An additional 648 women were excluded due to insufficient volumes of serum or urine samples. Of the remaining 1400 participants with serum samples available at the SWAN-MPS baseline, we excluded 232 women who had already reached natural menopause and 48 women who had had a hysterectomy and/or oophorectomy at the MPS baseline, resulting in a final analytic sample of 1120 women with 6586 observations and 5466 person-years of follow-up through 2017. Additional details of the study design are described elsewhere (47,48).

Ascertainment of natural menopause incidence

The age at the natural FMP was determined from annual interviews indicating 12 months of amenorrhea since the last menstrual period for no other causes (including hysterectomy, bilateral oophorectomy, or hormone therapy, HT). If a participant was reliably observed to have had a menstrual bleed followed by at least 12 consecutive months that were both HT free and bleed free, her FMP was ascertained. If a woman missed at least 3 consecutive visits prior to the first postmenopause visit, the FMP date was set to missing.

Measurements of PFAS serum concentrations

Baseline MPS serum samples were sent to the Division of Laboratory Sciences, National Center for Environmental Health, Centers for Disease Control and Prevention (CDC). The CDC laboratory's involvement did not constitute engagement in research of human subjects. Serum samples from subsequent SWAN visits were not analyzed because serum concentrations of the target analytes are relatively stable over time (48). We measured PFHxS, n-PFOS, sum of branched isomers of PFOS (Sm-PFOS), n-PFOA, sum of branched PFOA (Sb-PFOA), PFNA, perfluorodecanoic acid, perfluoroundecanoic acid, and perfluorododecanoic acid in 0.1 mL of serum, using an online solid phase extraction high-performance liquid chromatography isotope dilution tandem mass spectrometry method (49). The analytic methods and quality control procedures have been described elsewhere

(48). The coefficient of variation of low- and high-quality controls ranged from 6% to 12%. The limit of detection (LOD) was 0.1 ng/mL for all the analytes. Concentrations below the LODs were substituted with $\text{LOD}/\sqrt{2}$.

Assessments of covariates

Annual visits included an in-person interview, self-administered questionnaires, and measurements of weight and height. All questionnaires were translated into Spanish, Cantonese, and Japanese and back-translated; translation discrepancies were resolved by 2 translators.

Sociodemographic variables included race/ethnicity, study site, and educational attainment from the screening questionnaire. Race/ethnicity was classified into self-identified Black, Chinese, Japanese, or White. We categorized education as high school or less, some college, or college degree or higher. Baseline time-invariant health-related variables included prior oral contraceptive and other exogenous hormone use, and body mass index (BMI) at baseline. We did not consider time-varying BMI in case of overadjustment bias because PFAS might contribute to weight gain (50).

Time-varying lifestyle variables included annual self-reported active smoke exposure and physical activity. Self-reported smoking status was defined based on the questions asking about ever smoking, amount smoked, and quit date (51). Women were classified as never smoked, former smoked only, or current smoking. Physical activity was assessed using an adaptation of the Kaiser Physical Activity Survey (52), which consists of 38 questions with primarily Likert-scale responses about physical activity in various domains, including sports/exercise, household/caregiving, and daily routine (defined as walking or biking for transportation and hours of television watching, which are reverse coded). Domain-specific indices were derived by averaging the ordinal responses to questions in each domain, resulting in values from 1 to 5. Thus, the total physical activity score ranged from 3 to 15 with 15 indicating the highest level of activity.

Statistical analyses

Bivariate statistics were calculated for participant characteristics at baseline and PFAS serum concentrations stratified by racial/ethnic groups. Chi-square or Fisher's exact statistics were computed for categorical variables; and analysis of variance or Kruskal-Wallis tests were used for continuous variables. We censored a participant's data when she reported initiating HT if no subsequent HT-free bleeding occurred, at the date of hysterectomy or bilateral oophorectomy, or at the last menstrual period at the end of data collection if it occurred before 12 months of amenorrhea, on the date of death, or on the date of the participants' last follow-up visits. Of the 1120 participants, 578 had an observed date at the natural FMP. The remaining 542 were censored for one of the following reasons: hysterectomy and/or oophorectomy before having ≥ 12 months of amenorrhea ($n = 69$); had an unknown FMP date because of HT use ($n = 451$); or end of data collection before ≥ 12 months of amenorrhea ($n = 22$).

Hazard ratios (HRs) and 95% confidence intervals (CIs) of natural menopause incidence were estimated by Cox proportional hazards (PH) regression. We used time since baseline as the time scale. Serum PFAS concentrations were also categorized into tertiles. HRs and 95% CIs were calculated comparing

the medium and the highest tertiles of PFAS concentrations to the lowest tertiles (the reference group). To assess the linear trend of the associations between PFAS exposures and incident natural menopause, tertiles of PFAS concentrations were used as continuous variables in the regression models. We also tested the log-linear relationships using log-transformed PFAS concentrations (log-transformed with base 2). In this case, HRs and 95% CIs were interpreted as effects of a 2-fold increase in PFAS serum concentrations. Covariates considered in multivariate adjustments included baseline age (continuous), race/ethnicity (White, Black, Chinese, Japanese), educational attainment (high school or less, some college, college graduate, or post college), time-varying parity status (nulliparous, or parous), time-varying smoking status (never, former, or current smoker), time-varying physical activity, prior HT use at baseline, and baseline BMI. We applied the Cox PH models to generate adjusted survival curves and estimate median age at natural menopause. We calculated predicted survival probability of natural menopause (ie, probability of not having natural menopause). Median age at natural menopause was defined as the time at which 50% had reached their natural menopause. To examine effect modifications by race/ethnicity, we used statistical interaction terms between PFAS exposure and race/ethnicity. Chinese and Japanese were combined because of the small sample sizes of these groups.

People are exposed to multiple and often intercorrelated chemicals. Efforts to study chemical mixtures in isolation can thus result in underestimated environmental effects (53, 54). To identify subgroups corresponding to distinct MPS baseline PFAS concentration profiles, a nonparametric partitioning method, k-means clustering, was used to find an optimal number of clusters and assign membership to each cluster for each participant (55). The k-means clustering was conducted using PROC FASTCLUS procedures. All PFAS serum concentrations were log-transformed and standardized to z scores to make the distributions normal and comparable before the k-means analysis. The number of clusters was chosen based on cubic clustering criterion, pseudo F statistic (ie, the ratio of between-cluster variance to within-cluster variance), r-squared statistics, and interpretability. Participant characteristics differed significantly by k-means clusters (46). It is possible that there is uncertainty in classifying women based on their overall concentration patterns. Therefore, we utilized inverse probability treatment weighting method to account for confounding due to differences in distributions of these characteristics among clusters (56). HRs of natural menopause incidence were estimated for women with different clusters. We estimated conservative 95% CIs based on the robust variance estimator. All the analyses were performed using SAS, version 9.4 (SAS Institute, Inc., Cary, NC).

Sensitivity analyses

HT use or loss to follow up masked the actual FMP date. The SWAN Data Coordinating Center therefore conducted multiple imputations with chain equations for missing FMP age using a comprehensive list of covariates related to timing of menopause (see the list of covariates (46)). Covariates were selected based on previous literature (14, 15). The imputations were conducted using IVEware. Because we could not impute FMP age perfectly, we used 10 sets of imputations to account for uncertainty, and the pooled results were computed using PROC MIANALYZE.

Hysterectomy and/or oophorectomy was a competing risk in our analyses. Previous studies have suggested that exposures to PFOS, PFOA, and PFNA were associated with increased risk of endometriosis (57). Many women undergo a hysterectomy to help alleviate intolerable symptoms of endometriosis. Because such surgery would mask the age at which a woman would have become menopausal in the absence of surgery, the competing risk may preclude women from participation due to no longer being at risk of reaching the natural FMP. To examine the potential impact of this competing risk on our results, we excluded women who had hysterectomy in the sensitivity analyses. Furthermore, we excluded 29 women who reached their natural menopause since baseline to minimize the possibility of reverse causation bias. Lastly, it is possible that smokers enrolled in our study may be healthier than those who reached their natural menopause before baseline, especially those with premature menopause (before age 40) or early menopause (before age 45). We have conducted sensitivity analyses by removing smokers from our study sample.

Results

Study participants

The median (interquartile range, IQR) age of the 1120 premenopausal women was 48.9 (47.0–50.8) years with a range of 45 to 56 years at baseline (1999–2000) (Table 1). Most women had at least some college education. Educational attainment differed significantly by race/ethnicity, with Black women more likely to receive a high school education or less ($P < .0001$) than other racial/ethnic groups. Less than 40% of the women had ever smoked; a higher proportion of Black women were current smokers than the other racial/ethnic groups ($P < .0001$). Physical activity also differed significantly by race/ethnicity, with White women having higher activity scores ($P < .0001$). BMI at baseline was significantly higher among Black women and was the lowest in Chinese and Japanese women ($P < .0001$). Chinese and Japanese women were more likely to be nulliparous ($P < .0001$) and to report prior use of HT at baseline ($P = .0005$).

PFOS and PFOA were the PFASs detected at the highest concentrations (46). The median (IQR) serum concentration was 17.1 (12.2–24.5) ng/mL for n-PFOS, 7.2 (4.6–10.8) ng/mL for Sm-PFOS, and 4.0 (2.8–5.7) ng/mL for n-PFOA, 1.5 (0.9–2.3) ng/mL for PFHxS, and 0.6 (0.4–0.8) ng/mL for PFNA. Perfluoroundecanoic acid, perfluorododecanoic acid, perfluorodecanoic acid, and Sb-PFOA were detected in fewer than 40% of baseline samples, and thus they were not considered further in these analyses. Significant racial/ethnic differences were observed in serum PFAS concentrations: White women had the highest concentrations of n-PFOA; Black women had the highest concentrations of n-PFOS, and Sm-PFOS; Chinese and Japanese

Table 1. Baseline (1999-2000) characteristics of multi-racial/ethnic midlife women by racial/ethnic groups in the study of Women's Health Across the Nation (n = 1120)

Baseline characteristic	Total (n = 1120)	White (n = 577)	Black (n = 235)	Chinese (n = 142)	Japanese (n = 166)	P value ^a
	Median (IQR) or n (%)					
Age, years	48.9 (47.0-50.8)	48.7 (47.0-50.8)	48.7 (46.8-50.7)	49.3 (47.3-50.7)	49.2 (47.4-50.9)	.23
Study site						NA
Southeast MI	202 (18.0)	90 (15.6)	112 (47.7)	0	0	
Boston, MA	182 (16.3)	118 (20.4)	64 (27.2)	0	0	
Oakland, CA	242 (21.6)	100 (17.3)	0	142 (100)	0	
Los Angeles, CA	299 (26.7)	133 (23.1)	0	0	166 (100)	
Pittsburgh, PA	195 (17.4)	136 (23.6)	59 (25.1)	0	0	
Educational attainment						
≤High school	197 (17.7)	69 (12.0)	65 (28.0)	35 (24.7)	28 (16.9)	<.0001
Some college	350 (31.4)	174 (30.3)	90 (38.8)	28 (19.7)	58 (34.9)	
College	271 (24.3)	137 (23.9)	41 (17.7)	43 (30.3)	50 (30.1)	
Post college	296 (26.6)	194 (33.8)	36 (15.5)	36 (25.3)	30 (18.1)	
Parity						
Nulliparous	215 (19.2)	146 (25.3)	21 (8.9)	21 (14.8)	27 (16.3)	<.0001
Parous	905 (80.8)	431 (74.7)	214 (91.1)	121 (85.2)	139 (83.7)	
Prior hormone use	248 (22.1)	151 (26.2)	54 (23.0)	21 (14.8)	22 (13.3)	.0005
Smoking status						<.0001
Never smoker	720 (64.4)	343 (59.5)	134 (57.3)	134 (94.4)	109 (65.7)	
Former smoker	291 (26.0)	187 (32.5)	55 (23.5)	7 (4.9)	42 (25.3)	
Current smoker	107 (9.6)	46 (8.0)	45 (19.2)	1 (0.7)	15 (9.0)	
Physical activity score	7.9 (6.6-9.0)	8.1 (6.9-9.3)	7.3 (6.4-8.6)	7.2 (6.0-8.5)	7.8 (6.7-8.9)	<.0001
Body mass index, kg/m ²	26.1 (22.7-31.5)	26.5 (22.9-31.7)	31.4 (26.5-37.9)	23.0 (20.9-25.0)	23.3 (21.5-26.2)	<.0001

IQR, interquartile range. NA, not available.

^aChi-square tests or Fisher's exact tests were used for categorical variables; analysis of variance tests or Kruskal-Wallis tests were conducted for continuous variables. The significance level was set at .05.

women had the lowest PFHxS concentrations; White, Chinese, and Japanese women had a higher detection rate of PFNA, and significantly higher median concentrations than Black women. PFASs were positively correlated among each other with Spearman ρ s ranging from 0.35 to 0.82 (46).

Associations between PFAS and incident natural menopause

n-PFOS, Sm-PFOS, n-PFOA, and PFNA were associated with earlier age at natural FMP (Table 2). After multivariate adjustment for age at baseline, race/ethnicity, study site, education, parity, BMI at baseline, and time-varying physical activity and smoking status, and prior hormone use at baseline, comparing the highest with the lowest tertiles, the HR for natural menopause

was 1.26 (95% CI: 1.02-1.57) for n-PFOS ($P_{trend} = .03$), 1.27 (95% CI: 1.01-1.59) for Sm-PFOS ($P_{trend} = .03$), and 1.31 (95% CI: 1.04-1.65) for n-PFOA ($P_{trend} = .01$). The relationship between PFNA and incident natural menopause was not linear but log linear. The HR of natural menopause was 1.12 (95% CI: 1.01-1.24) per doubling increase in PFNA serum concentrations. No significant association with age of menopause was found for PFHxS in either trend ($P_{trend} = .24$) or log-linear analyses ($P = .15$). Adjusted survival curves by tertiles of PFAS concentrations are presented in (46). The predicted age at natural menopause in women with tertile 1, tertile 2, and tertile 3 of serum concentrations was 52.6 years, 52.3 years, and 51.6 years for n-PFOS; 52.6 years, 51.9 years, and 51.7 years for Sm-PFOS; 52.7 years, 51.9 years, and 51.6 years for n-PFOA;

Table 2. Hazard Ratios (HR) (95% Confidence Intervals, 95% CI) for Incident Natural Menopause with Tertile Changes and per Doubling Increase in Serum Concentrations of n-PFOS, Sm-PFOS, n-PFOA, PFNA, and PFHxS

PFAS	Tertile of PFAS concentrations			P value for trend ^c	Per doubling increase HR (95% CI)	P value ^c
	Tertile 1 HR (95% CI)	Tertile 2 HR (95% CI)	Tertile 3 HR (95% CI)			
n-PFOS						
Median (IQR), ng/mL	10.4 (8.1-12.2)	16.9 (15.6-18.7)	28.3 (24.2-37.8)			
no. cases/person-years	183/1861	192/1883	203/1880			
Model 1 ^a	Ref	1.04 (0.85-1.27)	1.19 (0.97-1.47)	.09	1.06 (0.96-1.18)	.26
Model 2 ^b	Ref	1.06 (0.86-1.31)	1.26 (1.02-1.57)	.03	1.11 (0.99-1.23)	.06
Sm-PFOS						
Median (IQR), ng/mL	3.8 (2.9-4.5)	7.1 (6.2-8.0)	13.0 (10.7-16.8)			
no. cases/person-years	195/1842	194/1923	189/1858			
Model 1 ^a	Ref	1.03 (0.84-1.27)	1.12 (0.90-1.39)	.30	1.04 (0.95-1.14)	.37
Model 2 ^b	Ref	1.11 (0.90-1.37)	1.27 (1.01-1.59)	.03	1.08 (0.99-1.19)	.09
n-PFOA						
Median (IQR), ng/mL	2.3 (1.8-2.8)	4.0 (3.5-4.5)	6.6 (5.6-8.6)			
No. cases/person-years	183/1818	195/1936	200/1870			
Model 1 ^a	Ref	1.15 (0.92-1.42)	1.29 (1.03-1.61)	.02	1.06 (0.95-1.19)	.27
Model 2 ^b	Ref	1.12 (0.90-1.40)	1.31 (1.04-1.65)	.01	1.11 (0.99-1.24)	.07
PFNA						
Median (IQR), ng/mL	0.3 (0.3-0.4)	0.5 (0.5-0.6)	0.9 (0.7-1.0)			
no. cases/person-years	168/1930	181/1679	229/2015			
Model 1 ^a	Ref	1.18 (0.95-1.46)	1.21 (0.99-1.49)	.07	1.13 (1.02-1.25)	.02
Model 2 ^b	Ref	1.18 (0.95-1.47)	1.20 (0.97-1.49)	.10	1.12 (1.01-1.24)	.04
PFHxS						
Median (IQR), ng/mL	0.8 (0.6-1.0)	1.5 (1.3-1.6)	3.0 (2.3-4.5)			
no. cases/person-years	203/1957	168/1728	207/1939			
Model 1 ^a	Ref	0.92 (0.75-1.13)	1.15 (0.94-1.41)	.19	1.04 (0.97-1.13)	.27
Model 2 ^b	Ref	1.05 (0.84-1.30)	1.11 (0.90-1.37)	.33	1.03 (0.95-1.11)	.50

^a Model 1 was adjusted for age at baseline, race/ethnicity, and study site.

^b Model 2 was additionally adjusted for education, parity, BMI at baseline, physical activity, smoking status, and prior hormone use at baseline.

^c The significance level was set at 0.05.

52.7 years, 51.8 years, and 51.8 years for PFNA; and 52.4 years, 51.9 years, and 51.8 years for PFHxS.

When we examined interaction terms between PFAS concentrations and race/ethnicity, significant associations with incidence of natural menopause were observed for PFNA and n-PFOA among White women but not in other racial/ethnic groups (Fig. 1). The HR for White women was 1.23 (95% CI: 1.06-1.44) and 1.33 (95% CI: 1.13-1.56) per doubling increase in serum concentrations of n-PFOA and PFNA, respectively, after covariate adjustment. The associations in Black or Asian women did not reach statistical significance. Neither did the results for n-PFOS, Sm-PFOS and PFHxS (46).

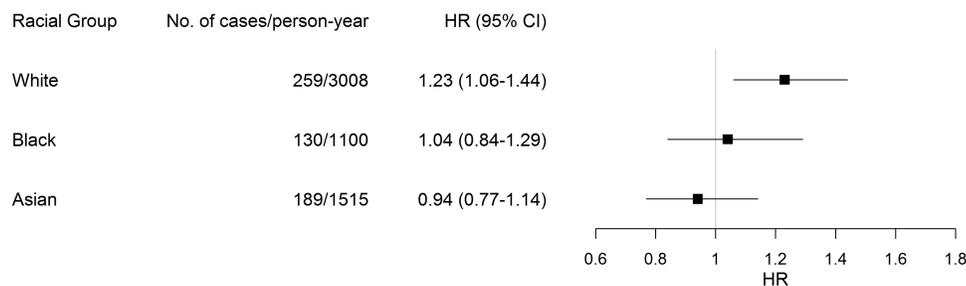
In sensitivity analyses, the pooled effect estimates from 10 imputations of age at FMP were largely unchanged, while the 95% CIs became narrower (Table 3). However, the significant associations between PFNA concentration and natural menopause disappeared. In the competing risks analyses, 67 women (303 observations) who had hysterectomy and/or oophorectomy were excluded from the analyses, but effect estimates remained similar (46). Exclusion of 29 women who reached natural menopause in the 6 months since baseline did not change results (46), diminishing the likelihood that reverse causation bias drove the observed

results. The results were robust when restricting the study sample to never smokers (46).

Mixture effects of PFAS on incident natural menopause

Participants were classified into clusters based on their overall PFAS concentrations profiles using the k-means method (46). Women were classified into 4 clusters based on serum PFAS concentrations, including “low,” “low–medium,” “medium–high,” and “high” overall concentration patterns. Women in the “low” concentration group had the lowest overall concentrations of PFAS, while those classified into the “high” group exhibited the highest concentrations. After adjusting for confounding, the HRs for natural menopause comparing the “high,” “medium–high,” “low–medium” concentration groups with the low concentration group were 1.63 (95% CI: 1.08-2.45), 1.31 (95% CI: 0.94-1.83), and 1.30 (95% CI: 0.97-1.74), respectively (46). Participants in the high concentration group had an earlier onset of natural menopause compared to those in other groups (Fig. 2). The predicted median age at natural menopause in the low concentration group was 52.8 years compared with 51.8 years, 52.0 years, and 50.8 years for low–medium, medium–high, and high concentration groups, respectively.

a) Exposure to n-PFOA and incidence of natural menopause by racial/ethnic groups



b) Exposure to PFNA and incidence of natural menopause by racial/ethnic groups

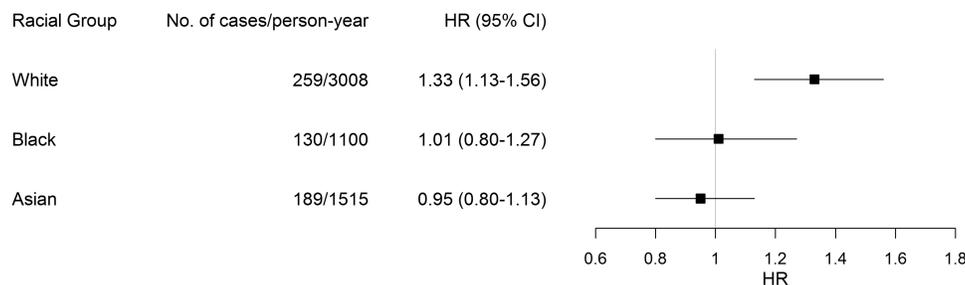


Figure 1. Adjusted hazard ratios (HR) (95% confidence intervals, 95% CI) for incident natural menopause with per doubling increase in serum concentrations of n-PFOA and PFNA. (A) Exposure to n-PFOA and incidence of natural menopause by racial/ethnic groups. (B) Exposure to PFNA and incidence of natural menopause by racial/ethnic groups. Models were adjusted for age at baseline, study site, education, parity, BMI at baseline, physical activity, smoking status, and prior hormone use at baseline. *P* values for the interaction terms with race/ethnicity are 0.08 for n-PFOA and 0.01 for PFNA.

Table 3. Pooled hazard ratios (HR) (95% confidence intervals, 95% CI) for incident natural menopause with tertile changes and per doubling increase in serum concentrations of n-PFOS, Sm-PFOS, n-PFOA, PFNA, and PFHxS with 10 imputations

PFAS	Tertile of PFAS concentrations				P value for trend ^d	Per doubling increase HR (95%CI)	P value ^d
	Tertile 1 HR (95%CI)	Tertile 2 HR (95%CI)	Tertile 3 HR (95%CI)				
n-PFOS							
Median (IQR), ng/mL	10.4 (8.1-12.2)	16.9 (15.6-18.7)	28.3 (24.2-37.8)				
No. cases/person-years ^a	315/1487	322/1499	344/1483				
Model 1 ^b	Ref	0.98 (0.84-1.16)	1.23 (1.05-1.46)	.01	1.10 (1.01-1.20)	.02	
Model 2 ^c	Ref	0.99 (0.84-1.17)	1.26 (1.06-1.49)	.01	1.11 (1.02-1.21)	.02	
Sm-PFOS							
Median (IQR), ng/mL	3.8 (2.9-4.6)	7.2 (6.2-8.1)	13.1 (10.9-17.2)				
No. cases/person-years ^a	320/1496	331/1510	330/1463				
Model 1 ^b	Ref	1.01 (0.86-1.19)	1.20 (1.01-1.43)	.04	1.09 (1.01-1.17)	.02	
Model 2 ^c	Ref	1.02 (0.86-1.20)	1.25 (1.04-1.50)	.01	1.11 (1.03-1.20)	.009	
n-PFOA							
Median (IQR), ng/mL	2.3 (1.8-2.8)	4.0 (3.5-4.5)	6.6 (5.6-8.6)				
No. cases/person-years ^a	313/1448	334/1553	334/1468				
Model 1 ^b	Ref	1.11 (0.94-1.30)	1.15 (0.98-1.35)	.06	1.10 (1.01-1.20)	.03	
Model 2 ^c	Ref	1.14 (0.96-1.35)	1.23 (1.03-1.47)	.02	1.10 (1.01-1.21)	.02	
PFNA							
Median (IQR), ng/mL	0.3 (0.3-0.4)	0.5 (0.5-0.6)	0.9 (0.7-1.0)				
No. cases/person-years ^a	331/1522	295/1362	374/1585				
Model 1 ^b	Ref	1.00 (0.85-1.19)	1.14 (0.96-1.34)	.12	1.07 (0.99-1.16)	.10	
Model 2 ^c	Ref	0.98 (0.82-1.18)	1.11 (0.94-1.33)	.20	1.05 (0.97-1.14)	.23	
PFHxS							
Median (IQR), ng/mL	0.8 (0.6-1.0)	1.5 (1.3-1.6)	3.0 (2.3-4.5)				
No. cases/person-years ^a	337/1592	299/1324	344/1553				
Model 1 ^b	Ref	1.08 (0.90-1.28)	1.13 (0.97-1.35)	.10	1.05 (0.99-1.12)	.09	
Model 2 ^c	Ref	1.02 (0.86-1.23)	1.11 (0.94-1.31)	.24	1.05 (0.98-1.12)	.15	

^aAveraged no. cases and person-years from 10 imputations.^bModel 1 was adjusted for age at baseline, race/ethnicity, and study site.^cModel 2 was additionally adjusted for education, parity, BMI at baseline, physical activity, smoking status, and prior hormone use at baseline.^dThe significance level was set at .05.

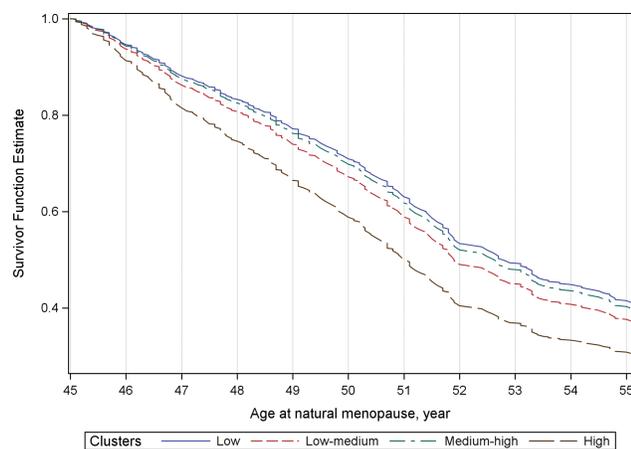


Figure 2. Adjusted survival curves for natural menopause by participant clusters. The model was adjusted for age at baseline, race/ethnicity, study site, education, parity, BMI at baseline, physical activity, smoking status, and prior hormone use at baseline. The hazards ratio for low–medium, medium–high, and high groups were 1.30 (95% CI: 0.97–1.74), 1.31 (95% CI: 0.94–1.83), and 1.63 (95% CI: 1.08–2.45), respectively, compared to the low group. The predicted median age at natural menopause for women with low overall PFAS concentration profile was 52.8 years, and 51.8 years, 52.0 years and 50.8 years for those with low–medium, medium–high, and high overall concentration patterns, respectively.

Discussion

In this 17-year prospective cohort of 1120 women with 5466 person-years of observation in annual follow-up visits, we found that higher baseline serum concentrations of n-PFOS, Sm-PFOS, n-PFOA, and PFNA were significantly associated with an earlier age at natural FMP. PFHxS concentrations were not associated with incidence of natural menopause. The analysis of mixtures also suggested that the combined PFAS mixtures were associated with earlier onset of natural menopause. These results suggest that PFAS may play an important role in ovarian aging, perhaps through its endocrine disruptive actions.

Comparison with previous epidemiologic studies

To date, evidence on the influence of PFAS exposure on the timing of menopause and ovarian aging has been limited and inconsistent, and has been primarily generated from cross-sectional studies that could not establish causal relationships (38, 39). Knox et al. found that higher concentrations of PFOA and PFOS were associated with earlier menopausal age in a cross-sectional study of women aged 18 to 65 years from the C8 Health Project (38). This study collected data from highly exposed communities and workers in six public water districts contaminated with PFOA from the DuPont Washington Works Plant near Parkersburg (58). Taylor et al. reported significant relationships between higher

PFHxS concentrations and earlier menopause, but not for PFOA, PFOS, and PFNA among the US general women aged 20 to 65 years from NHANES (39). Using estimated retrospective year-specific serum concentrations for 1951–2011 and PFOA concentrations measured in 2005–2006, no association was observed between earlier age at menopause with PFOA exposure in either retrospective or prospective cohort of C8 Science Panel (43). However, reverse causation could not be ruled out as women appeared to have higher PFAS concentrations after menopause (39–42).

Serum concentrations of PFAS appear to be higher in males than in females across all age groups (59). Wong et al (60). found that menstruation could explain the PFOS elimination half-life difference between men and women. The differences by sex narrows with age, suggesting that PFAS may reaccumulate after cessation of menstrual bleeding in postmenopausal women (41, 42, 47, 48). Given that 90% to 99% of PFASs in the blood are bound to serum albumin (61, 62), menstrual bleeding could be an important elimination pathway in women. It is still unclear whether PFAS exposure is the cause of earlier natural menopause or the cessation of menstruation leads to increased serum concentrations of PFAS. Therefore, previously observed associations identified in cross-sectional or retrospective designs (38, 39, 43) could result from the impact of reproductive aging on serum PFAS concentrations, rather than their adverse effects on ovarian reserve.

To our knowledge, the current investigation is among the first of studies to evaluate the associations of exposures to various PFAS with the occurrence of natural menopause in a prospective cohort of multiracial/ethnic midlife women. Our findings of PFOA and timing of natural menopause are not in concordance with Dhingra et al. (43), the only other published study to our knowledge that has explored the associations between PFOA exposure and incident menopause. Dhingra et al. (43) included 8759 women aged ≥ 40 years in 2005–2006 who were exposed to high levels of PFOA from a chemical facility in the Mid-Ohio Valley, West Virginia (the C8 Health Project). Women were recruited in 2005–2006 and interviewed in 2008–2011 to ascertain the timing of menopause. They found no significant association between PFOA exposure (using either estimated cumulative exposure during 1951 and 2011, or measured serum concentrations in 2005–2006) and incident natural menopause. They observed that women exhibited considerable digit preference and tended to round off their age at menopause to 40, 45, or 55, suggesting that their results may suffer from the reduced accuracy

of recalled ages and are presumably biased towards the null. The availability of a standardized questionnaire administered prospectively at regular, approximately annual, intervals to confirm menopausal status and ascertain age at the natural FMP is a major strength of SWAN and may account for the observed differences in the findings.

No previous research of which we are aware has explored the mixture effects of PFAS on ovarian aging. PFAS are ubiquitous and environmentally persistent (63). People may be normally exposed to multiple PFAS through drinking water, food intake, or use of consumer products (64, 65). Understanding concentration patterns of multiple PFAS is an important first step before examining the association between PFAS mixtures and incident natural menopause. Results of mixture analyses showed a larger joint effect on ovarian aging compared with single PFAS. Along with our recent study of profiles of urinary concentrations of metal mixtures among midlife women (66), the results of this study suggested that k-means clustering is a useful tool to identify clusters in the population.

This is also the first study of which we are aware to explore effect modification by race/ethnicity on the associations between PFAS exposure and natural menopause. Although environmental exposure in general is sometimes expected to be higher in racial minority groups and socioeconomically disadvantaged neighborhoods, the concentration patterns tended to depend on the PFASs. Serum concentrations of n-PFOA were found to be higher in White women and PFNA concentrations were relatively higher in White and Chinese women, whereas serum concentrations of n-PFOS and Sm-PFOS were higher in Black women. This is consistent with previous findings (47, 67-69). White women with higher n-PFOA and PFNA tended to have earlier natural menopause.

PFOA is used as a surfactant and emulsifier in compounds used to coat a variety of food packaging materials, including microwave popcorn bags (21-23) and is essential in manufacture of the fluoropolymer polytetrafluoroethylene used in nonstick coatings and waterproof fabrics (70). Uses of PFOS included inks, varnishes, waxes, fire-fighting foams, and coating formulations (71). Use of consumer products may have contributed to more exposure to PFOA, while the most dominant source of PFOS exposure might have been intake of contaminated drinking water (28). Although production and use of some PFAS, including PFOA and PFOS, in the United States is on the decline, environmental exposures to many of these pervasive chemicals continue with associated potential hazards to human reproductive health.

Our study results showed no difference in the effects of n-PFOS and Sm-PFOS by racial/ethnic groups, possibly because of the exclusion of women with premature (before the age of 40 years) or early menopause (before age 45 years), or greater censoring of Black women who had surgical menopause before age 45. However, caution should be taken in interpreting the findings because of the modest sample sizes in those racial/ethnic groups. Asian women with similar PFNA concentrations as White women did not reach their natural menopause earlier. Previous studies have shown increases in PFNA concentrations since 2000 (59, 67, 72-74). Future studies with more recent PFNA measurements are needed to confirm our findings and better understand exposure trends. It is also important to explore the role of genetic background and changes in lifestyle factors (75-78).

Biologic evidence

PFAS exposures have been associated with diminished ovarian reserve (ie, the number of ovarian follicles and oocytes) (79-86). The mechanisms of PFAS-induced effects have widely been thought to occur through a PPAR mechanism (35, 36, 87). PPARs are expressed in the female hypothalamic-pituitary-gonadal axis, and they act on critical processes for ovarian function. For example, PPARs may inhibit transactivation of the estrogen receptor (ER) through competition for estrogen response element binding (88), down-regulate aromatase expression via nuclear factor- κ B pathway (89), and affect enzymatic activity in steroidogenesis (56, 90).

Accumulating evidence from experimental research suggests that PFAS can directly interfere with steroidogenic enzyme activities (33, 91, 92). Recently, it was also reported that PFNA and PFOA are weak xenoestrogens, inducing ER α -dependent transcriptional activation *in vitro* and *in vivo* (93). As potential endocrine disruptors, PFAS might also suppress the effects of 17 β -estradiol on estrogen-responsive gene expression (91, 94), reduce 17 β -estradiol production and alter the expression of major steroidogenic genes and regulator steroidogenic factors 1 (95). Disruption of ER signaling pathways may contribute to adverse health effects, such as reproductive failure and acceleration of ovarian aging, thus supporting the notion that women may be particularly vulnerable to reproductive toxicity of PFAS. In addition, experimental studies suggest that PFOA may lead to minimal but significant histopathologic changes in the uterus, vagina, and cervix (32).

Strengths and limitations

The primary strengths of this study included direct measurements of PFAS serum concentrations prior to

menopause, prospectively determination of FMP date, and a large cohort of community-based midlife women from 4 racial/ethnic groups followed for up to 17 years. The reproductive toxicity of PFAS has not been previously characterized among Chinese and Japanese women, to our knowledge. The prospective design also minimized the possibility of reverse causation. Standard annual follow-up visits instead of one-time questionnaire provided reliable estimates of date of FMP. We also consider multiple factors simultaneously in the Cox PH model, censoring at initiation of HT use or at hysterectomy or oophorectomy, thus providing HRs for natural menopause for the independent relations of all exposure factors examined.

Several limitations should be considered as well. First, enrollment at age 45 to 56 years was limited to menstruating women, thus women with earlier menopause were excluded. This left truncation resulted in an overestimation of median age at FMP (96). Women who experienced menopause before baseline, especially those with premature menopause (before age 40 years) or early menopause (before age 45 years), were not included in the cohort, which could bias our effect estimates towards the null. However, the effect estimates remained similar when restricting our study sample to never smokers. Second, more than 40% of the cohort was censored at the initiation of HT, before the participants were classified as postmenopausal. This could have resulted in an underestimation of the age at FMP because these women had higher education levels, which has been associated with later age at menopause. To minimize potential bias, we imputed their FMP age based on covariates related to the timing of menopause. Imputing age at menopause increased sample size and broadened generalizability to women with HT use and thus might have reduced bias. Finally, hysterectomy could be a competing risk of natural menopause. Hysterectomy can be undertaken for medical conditions (such as endometriosis or uterine fibroids, cancer or menorrhagia). We did not have data on the date of onset of these conditions and hence were unable to examine directly the potential effects of PFAS on cause-specific subsets of menopause (either surgically or naturally occurring).

Conclusions

Our findings suggest that exposure to select PFAS was associated with earlier natural menopause. Women with highest tertiles of n-PFOS serum concentrations tended to have 1.0 years earlier median time to natural menopause, and 0.9 years and 1.1 years earlier for Sm-PFOS and n-PFOA, respectively, compared with those in the lowest tertiles. High overall PFAS concentration patterns might contribute to 2.0 years earlier median time

to natural menopause compared with the low group. These estimates were roughly equivalent to or even larger than an effect estimate of 1.1 years comparing current smokers versus never smokers in our sample. Due to PFAS widespread use and environmental persistence, their potential adverse effects remain a public health concern.

Acknowledgments

We thank the study staff at each site and all the women who participated in SWAN.

We also thank the late Ms. Xiaoyun Ye at CDC for her support in PFAS assessment.

Financial Support: The Study of Women's Health Across the Nation (SWAN) has grant support from the National Institutes of Health (NIH), DHHS, through the National Institute on Aging (NIA), the National Institute of Nursing Research (NINR) and the NIH Office of Research on Women's Health (ORWH) (Grants U01NR004061; U01AG012505, U01AG012535, U01AG012531, U01AG012539, U01AG012546, U01AG012553, U01AG012554, U01AG012495). The study was also supported by the SWAN Repository (U01AG017719). This publication was supported in part by the National Center for Research Resources and the National Center for Advancing Translational Sciences, National Institutes of Health, through UCSF-CTSI Grant Number UL1 RR024131. This study was also supported by grants from the National Institute of Environmental Health Sciences (NIEHS) R01-ES026578, R01-ES026964 and P30-ES017885, and by the Centers for Disease Control and Prevention (CDC)/National Institute for Occupational Safety and Health (NIOSH) grant T42-OH008455.

Additional Information

Correspondence and Reprint Requests: Sung Kyun Park, Department of Epidemiology, University of Michigan, M5541 SPH II, 1415 Washington Heights, Ann Arbor, MI 48109-2029. E-mail: sungkyun@umich.edu.

Disclosure Summary: The authors declare no competing financial interest. All authors have submitted the ICMJE Form for Disclosure of Potential Conflicts of Interest. Conflicts that the editors consider relevant to the content of the manuscript have been disclosed. The content of this article is solely the responsibility of the authors and does not necessarily represent the official views of the NIA, NINR, ORWH, or the NIH. The findings and conclusions of this report are those of the authors and do not necessarily represent the official position of the Centers for Disease Control and Prevention (CDC). Use of trade names is for identification only and does not imply endorsement by the CDC, the Public Health Service, or the US Department of Health and Human Services. **Clinical Centers:** University of Michigan, Ann Arbor – Siobán Harlow, PI 2011-present, MaryFran Sowers, PI 1994-2011; Massachusetts General Hospital, Boston, MA – Joel

Finkelstein, PI 1999-present; Robert Neer, PI 1994-1999; Rush University, Rush University Medical Center, Chicago, IL – Howard Kravitz, PI 2009-present; Lynda Powell, PI 1994-2009; University of California, Davis/Kaiser – Ellen Gold, PI; University of California, Los Angeles – Gail Greendale, PI; Albert Einstein College of Medicine, Bronx, NY – Carol Derby, PI 2011-present, Rachel Wildman, PI 2010-2011; Nanette Santoro, PI 2004-2010; University of Medicine and Dentistry – New Jersey Medical School, Newark – Gerson Weiss, PI 1994-2004; and the University of Pittsburgh, Pittsburgh, PA-Karen Matthews, PI. *NIH Program Office*: National Institute on Aging, Bethesda, MD – Chhanda Dutta 2016-present; Winifred Rossi 2012-2016; Sherry Sherman 1994-2012; Marcia Ory 1994-2001; National Institute of Nursing Research, Bethesda, MD – Program Officers. *Central Laboratory*: University of Michigan, Ann Arbor – Daniel McConnell (Central Ligand Assay Satellite Services). *SWAN Repository*: University of Michigan, Ann Arbor – Siobán Harlow 2013-present; Dan McConnell 2011-2013; MaryFran Sowers 2000-2011. *Coordinating Center*: University of Pittsburgh, Pittsburgh, PA – Maria Mori Brooks, PI 2012-present; Kim Sutton-Tyrrell, PI 2001-2012; New England Research Institutes, Watertown, MA – Sonja McKinlay, PI 1995-2001. Steering Committee: Susan Johnson, Current Chair; Chris Gallagher, Former Chair.

Data Availability: Restrictions apply to the availability of data generated or analyzed during this study to preserve patient confidentiality or because they were used under license. The corresponding author will on request detail the restrictions and any conditions under which access to some data may be provided.

References

- Snowdon DA, Kane RL, Beeson WL, et al. Is early natural menopause a biologic marker of health and aging? *Am J Public Health*. 1989;79(6):709-714.
- Wise PM, Krajnak KM, Kashon ML. Menopause: the aging of multiple pacemakers. *Science*. 1996;273(5271):67-70.
- Mondul AM, Rodriguez C, Jacobs EJ, Calle EE. Age at natural menopause and cause-specific mortality. *Am J Epidemiol*. 2005;162(11):1089-1097.
- Ossewaarde ME, Bots ML, Verbeek AL, et al. Age at menopause, cause-specific mortality and total life expectancy. *Epidemiology*. 2005;16(4):556-562.
- Jacobsen BK, Heuch I, Kvåle G. Age at natural menopause and all-cause mortality: a 37-year follow-up of 19,731 Norwegian women. *Am J Epidemiol*. 2003;157(10):923-929.
- Atsma F, Bartelink ML, Grobbee DE, van der Schouw YT. Postmenopausal status and early menopause as independent risk factors for cardiovascular disease: a meta-analysis. *Menopause*. 2006;13(2):265-279.
- Hu FB, Grodstein F, Hennekens CH, et al. Age at natural menopause and risk of cardiovascular disease. *Arch Intern Med*. 1999;159(10):1061-1066.
- van der Schouw YT, van der Graaf Y, Steyerberg EW, Eijkemans JC, Banga JD. Age at menopause as a risk factor for cardiovascular mortality. *Lancet*. 1996;347(9003):714-718.
- de Kleijn MJ, van der Schouw YT, Verbeek AL, Peeters PH, Banga JD, van der Graaf Y. Endogenous estrogen exposure and cardiovascular mortality risk in postmenopausal women. *Am J Epidemiol*. 2002;155(4):339-345.
- Parazzini F, Bidoli E, Franceschi S, et al. Menopause, menstrual and reproductive history, and bone density in northern Italy. *J Epidemiol Community Health*. 1996;50(5):519-523.
- Kritz-Silverstein D, Barrett-Connor E. Early menopause, number of reproductive years, and bone mineral density in postmenopausal women. *Am J Public Health*. 1993;83(7):983-988.
- Shuster LT, Rhodes DJ, Gostout BS, Grossardt BR, Rocca WA. Premature menopause or early menopause: long-term health consequences. *Maturitas*. 2010;65(2):161-166.
- de Bruin JP, Bovenhuis H, van Noord PA, et al. The role of genetic factors in age at natural menopause. *Hum Reprod*. 2001;16(9):2014-2018.
- Gold EB, Bromberger J, Crawford S, et al. Factors associated with age at natural menopause in a multiethnic sample of midlife women. *Am J Epidemiol*. 2001;153(9):865-874.
- Gold EB, Crawford SL, Avis NE, et al. Factors related to age at natural menopause: longitudinal analyses from SWAN. *Am J Epidemiol*. 2013;178(1):70-83.
- Vabre P, Gatimel N, Moreau J, et al. Environmental pollutants, a possible etiology for premature ovarian insufficiency: a narrative review of animal and human data. *Environ Health*. 2017;16(1):37.
- Grindler NM, Allsworth JE, Macones GA, Kannan K, Roehl KA, Cooper AR. Persistent organic pollutants and early menopause in U.S. women. *PLoS One*. 2015;10(1):e0116057.
- Diamanti-Kandarakis E, Bourguignon JP, Giudice LC, et al. Endocrine-disrupting chemicals: an Endocrine Society scientific statement. *Endocr Rev*. 2009;30(4):293-342.
- Bradley EL, Read WA, Castle L. Investigation into the migration potential of coating materials from cookware products. *Food Addit Contam*. 2007;24(3):326-335.
- Sinclair E, Kim SK, Akinleye HB and, Kannan K. Quantitation of gas-phase perfluoroalkyl surfactants and fluorotelomer alcohols released from nonstick cookware and microwave popcorn bags. *Environ Sci Technol*. 2007;41(4):1180-1185.
- Trier X, Granby K, Christensen JH. Polyfluorinated surfactants (PFS) in paper and board coatings for food packaging. *Environ Sci Pollut Res Int*. 2011;18(7):1108-1120.
- Schaider LA, Balan SA, Blum A, et al. Fluorinated Compounds in U.S. Fast Food Packaging. *Environ Sci Technol Lett*. 2017;4(3):105-111.
- Begley TH, White K, Honigfort P, Twaroski ML, Neches R, Walker RA. Perfluorochemicals: potential sources of and migration from food packaging. *Food Addit Contam*. 2005;22(10):1023-1031.
- Hill PJ, Taylor M, Goswami P, Blackburn RS. Substitution of PFAS chemistry in outdoor apparel and the impact on repellency performance. *Chemosphere*. 2017;181:500-507.
- Lee JH, Lee CK, Suh CH, Kang HS, Hong CP, Choi SN. Serum concentrations of per- and poly-fluoroalkyl substances and factors associated with exposure in the general adult population in South Korea. *Int J Hyg Environ Health*. 2017;220(6):1046-1054.
- Kantiani L, Llorca M, Sanchis J, Farré M, Barceló D. Emerging food contaminants: a review. *Anal Bioanal Chem*. 2010;398(6):2413-2427.
- Butenhoff JL, Olsen GW, Pfahles-Hutchens A. The applicability of biomonitoring data for perfluorooctanesulfonate to the environmental public health continuum. *Environ Health Perspect*. 2006;114(11):1776-1782.
- Trudel D, Horowitz L, Wormuth M, Scheringer M, Cousins IT, Hungerbühler K. Estimating consumer exposure to PFOS and PFOA. *Risk Anal*. 2008;28(2):251-269.
- Kissa E. *Fluorinated Surfactants and Repellents*. CRC Press; 2001.
- Ding N, Harlow SD, Randolph Jr JF, Loch-Carusso R, Park SK. Perfluoroalkyl and polyfluoroalkyl substances (PFAS) and their effects on the ovary. *Hum Reprod Update*. 2020:dmaa018. doi:10.1093/humupd/dmaa018

31. Kar S, Sepúlveda MS, Roy K, Leszczynski J. Endocrine-disrupting activity of per- and polyfluoroalkyl substances: exploring combined approaches of ligand and structure based modeling. *Chemosphere*. 2017;184:514-523.
32. Dixon D, Reed CE, Moore AB, et al. Histopathologic changes in the uterus, cervix and vagina of immature CD-1 mice exposed to low doses of perfluorooctanoic acid (PFOA) in a uterotrophic assay. *Reprod Toxicol*. 2012;33(4):506-512.
33. Chaparro-Ortega A, Betancourt M, Rosas P, et al. Endocrine disruptor effect of perfluorooctane sulfonic acid (PFOS) and perfluorooctanoic acid (PFOA) on porcine ovarian cell steroidogenesis. *Toxicol in Vitro*. 2018;46:86-93.
34. Zhao Y, Tan YS, Strynner MJ, Perez G, Haslam SZ, Yang C. Perfluorooctanoic acid effects on ovaries mediate its inhibition of peripubertal mammary gland development in Balb/c and C57Bl/6 mice. *Reprod Toxicol*. 2012;33(4):563-576.
35. Andersen ME, Butenhoff JL, Chang SC, et al. Perfluoroalkyl acids and related chemistries—toxicokinetics and modes of action. *Toxicol Sci*. 2008;102(1):3-14.
36. White SS, Fenton SE, Hines EP. Endocrine disrupting properties of perfluorooctanoic acid. *J Steroid Biochem Mol Biol*. 2011;127(1-2):16-26.
37. Lau C, Anitole K, Hodes C, Lai D, Pfahles-Hutchens A, Seed J. Perfluoroalkyl acids: a review of monitoring and toxicological findings. *Toxicol Sci*. 2007;99(2):366-394.
38. Knox SS, Jackson T, Javins B, Frisbee SJ, Shankar A, Ducatman AM. Implications of early menopause in women exposed to perfluorocarbons. *J Clin Endocrinol Metab*. 2011;96(6):1747-1753.
39. Taylor KW, Hoffman K, Thayer KA, Daniels JL. Polyfluoroalkyl chemicals and menopause among women 20-65 years of age (NHANES). *Environ Health Perspect*. 2014;122(2):145-150.
40. Konkel L. PFCs and early menopause: association raises questions about causality. *Environ Health Perspect*. 2014;122(2):A59.
41. Dhingra R, Winkvist A, Darrow LA, Klein M, Steenland K. A study of reverse causation: examining the associations of perfluorooctanoic acid serum levels with two outcomes. *Environ Health Perspect*. 2017;125(3):416-421.
42. Ruark CD, Song G, Yoon M, et al. Quantitative bias analysis for epidemiological associations of perfluoroalkyl substance serum concentrations and early onset of menopause. *Environ Int*. 2017;99:245-254.
43. Dhingra R, Darrow LA, Klein M, Winkvist A, Steenland K. Perfluorooctanoic acid exposure and natural menopause: a longitudinal study in a community cohort. *Environ Res*. 2016;146:323-330.
44. Santoro N, Johnson J. Diagnosing the onset of menopause. *JAMA*. 2019;322(8):775-776.
45. Sowers MF, Crawford S, Sternfeld B, et al. SWAN: a multicenter, multiethnic, community-based cohort study of women and the menopausal transition. *Women's Heal. Res. Fac. Publ*. 2000. Available at: https://escholarship.umassmed.edu/wfc_pp/505. Accessed July 4, 2018.
46. Ding N, Harlow SD, Randolph JF, Jr., et al. 2020. Associations between Per- and Polyfluoroalkyl Substances and Incident Natural Menopause, Github, Supplemental Data. https://github.com/um-mpeg/Publications_2
47. Park SK, Peng Q, Ding N, Mukherjee B, Harlow SD. Determinants of per- and polyfluoroalkyl substances (PFAS) in midlife women: evidence of racial/ethnic and geographic differences in PFAS exposure. *Environ Res*. 2019;175:186-199.
48. Ding N, Harlow SD, Batterman S, Mukherjee B, Park SK. Longitudinal trends in perfluoroalkyl and polyfluoroalkyl substances among multiethnic midlife women from 1999 to 2011: the Study of Women's Health Across the Nation. *Environ Int*. 2020;135:105381.
49. Kato K, Basden BJ, Needham LL, Calafat AM. Improved selectivity for the analysis of maternal serum and cord serum for polyfluoroalkyl chemicals. *J Chromatogr A*. 2011;1218(15):2133-2137.
50. Liu G, Dhana K, Furtado JD, et al. Perfluoroalkyl substances and changes in body weight and resting metabolic rate in response to weight-loss diets: a prospective study. *PLoS Med*. 2018;15(2):e1002502.
51. Ferris BG. Epidemiology Standardization Project (American Thoracic Society). *Am Rev Respir Dis*. 1978;118(6 Pt 2):1-120.
52. Sternfeld B, Ainsworth BE, Quesenberry CP. Physical activity patterns in a diverse population of women. *Prev Med*. 1999;28(3):313-323.
53. Wang X, Mukherjee B, Park SK. Associations of cumulative exposure to heavy metal mixtures with obesity and its comorbidities among U.S. adults in NHANES 2003–2014. *Environ. Int*. 2018;121:683-694.
54. Wang X, Mukherjee B, Park SK. Does information on blood heavy metals improve cardiovascular mortality prediction? *J Am Heart Assoc*. 2019;8(21):e013571.
55. Jain AK. Data clustering: 50 years beyond K-means. *Pattern Recognit. Lett*. 2010;31(8):651-666.
56. Toda K, Okada T, Miyaura C, Saibara T. Fenofibrate, a ligand for PPARalpha, inhibits aromatase cytochrome P450 expression in the ovary of mouse. *J Lipid Res*. 2003;44(2):265-270.
57. Campbell S, Raza M, Pollack AZ. Perfluoroalkyl substances and endometriosis in US women in NHANES 2003-2006. *Reprod Toxicol*. 2016;65:230-235.
58. Frisbee SJ, Brooks AP Jr, Maher A, et al. The C8 health project: design, methods, and participants. *Environ Health Perspect*. 2009;117(12):1873-1882.
59. Calafat AM, Wong LY, Kuklennyik Z, Reidy JA, Needham LL. Polyfluoroalkyl chemicals in the U.S. population: data from the National Health and Nutrition Examination Survey (NHANES) 2003-2004 and comparisons with NHANES 1999-2000. *Environ Health Perspect*. 2007;115(11):1596-1602.
60. Wong F, MacLeod M, Mueller JF, Cousins IT. Enhanced elimination of perfluorooctane sulfonic acid by menstruating women: evidence from population-based pharmacokinetic modeling. *Environ Sci Technol*. 2014;48(15):8807-8814.
61. Han X, Snow TA, Kemper RA, Jepson GW. Binding of perfluorooctanoic acid to rat and human plasma proteins. *Chem Res Toxicol*. 2003;16(6):775-781.
62. Ylisen M, Auriola S. Tissue distribution and elimination of perfluorodecanoic acid in the rat after single intraperitoneal administration. *Pharmacol Toxicol*. 1990;66(1):45-48.
63. Olsen GW, Burris JM, Ehresman DJ, et al. Half-life of serum elimination of perfluorooctanesulfonate, perfluorohexanesulfonate, and perfluorooctanoate in retired fluorochemical production workers. *Environ Health Perspect*. 2007;115(9):1298-1305.
64. Domingo JL, Nadal M. Per- and Polyfluoroalkyl Substances (PFASs) in food and human dietary intake: a review of the recent scientific literature. *J Agric Food Chem*. 2017;65(3):533-543.
65. Domingo JL, Nadal M. Human exposure to per- and polyfluoroalkyl substances (PFAS) through drinking water: a review of the recent scientific literature. *Environ Res*. 2019;177:108648.
66. Wang X, Mukherjee B, Batterman S, Harlow SD, Park SK. Urinary metals and metal mixtures in midlife women: the Study of Women's Health Across the Nation (SWAN). *Int J Hyg Environ Health*. 2019;222(5):778-789.
67. Calafat AM, Kuklennyik Z, Reidy JA, Caudill SP, Tully JS, Needham LL. Serum concentrations of 11 polyfluoroalkyl compounds in the U.S. population: data from the national health and nutrition examination survey (NHANES). *Environ Sci Technol*. 2007;41(7):2237-2242.
68. Jain RB. Contribution of diet and other factors to the levels of selected polyfluorinated compounds: data from NHANES 2003–2008. *Int. J. Hyg. Environ. Health*. 2014;217(1):52-61.

69. Lindstrom AB, Strynar MJ, Libelo EL. Polyfluorinated compounds: past, present, and future. *Environ Sci Technol*. 2011;45(19):7954-7961.
70. Paul AG, Jones KC, Sweetman AJ. A first global production, emission, and environmental inventory for perfluorooctane sulfonate. *Environ Sci Technol*. 2009;43(2):386-392.
71. EFSA. Perfluorooctane sulfonate (PFOS), perfluorooctanoic acid (PFOA) and their salts. Scientific Opinion of the Panel on Contaminants in the Food chain. *EFSA J*. 2008;653:1-131.
72. Calafat AM, Kuklenyik Z, Caudill SP, Reidy JA, Needham LL. Perfluorochemicals in pooled serum samples from United States residents in 2001 and 2002. *Environ Sci Technol*. 2006;40(7):2128-2134.
73. Kato K, Wong LY, Jia LT, Kuklenyik Z, Calafat AM. Trends in exposure to polyfluoroalkyl chemicals in the U.S. Population: 1999-2008. *Environ Sci Technol*. 2011;45(19):8037-8045.
74. Spliethoff HM, Tao L, Shaver SM, et al. Use of newborn screening program blood spots for exposure assessment: declining levels of perfluorinated compounds in New York State infants. *Environ Sci Technol*. 2008;42(14):5361-5367.
75. Ding N, Wang X, Weisskopf MG, et al. Lead-related genetic loci, cumulative lead exposure and incident coronary heart disease: the normative aging study. *PLoS One*. 2016;11(9):e0161472.
76. Wang X, Ding N, Tucker KL, et al. A western diet pattern is associated with higher concentrations of blood and bone lead among middle-aged and elderly men. *J Nutr*. 2017;147(7):1374-1383.
77. Ding N, Wang X, Tucker KL, et al. Dietary patterns, bone lead and incident coronary heart disease among middle-aged to elderly men. *Environ Res*. 2019;168:222-229.
78. Wang X, Kim D, Tucker KL, Weisskopf MG, Sparrow D, Hu H, Park SK. Effect of dietary sodium and potassium intake on the mobilization of bone lead among middle-aged and older men: the veterans affairs normative aging study. *Nutrients*. 2019;11(11):2750. doi: [10.3390/nu11112750](https://doi.org/10.3390/nu11112750)
79. Du G, Hu J, Huang Z, et al. Neonatal and juvenile exposure to perfluorooctanoate (PFOA) and perfluorooctane sulfonate (PFOS): advance puberty onset and kisspeptin system disturbance in female rats. *Ecotoxicol Environ Saf*. 2019;167:412-421.
80. Chen Y, Zhou L, Xu J, et al. Maternal exposure to perfluorooctanoic acid inhibits luteal function via oxidative stress and apoptosis in pregnant mice. *Reprod Toxicol*. 2017;69:159-166.
81. Feng X, Wang X, Cao X, Xia Y, Zhou R, Chen L. Chronic exposure of female mice to an environmental level of perfluorooctane sulfonate suppresses estrogen synthesis through reduced histone H3K14 acetylation of the StAR promoter leading to deficits in follicular development and ovulation. *Toxicol Sci*. 2015;148(2):368-379.
82. Bellingham M, Fowler PA, Amezaga MR, et al. Exposure to a complex cocktail of environmental endocrine-disrupting compounds disturbs the kisspeptin/GPR54 system in ovine hypothalamus and pituitary gland. *Environ Health Perspect*. 2009;117(10):1556-1562.
83. Feng X, Cao X, Zhao S, et al. Exposure of pregnant mice to perfluorobutanesulfonate causes hypothyroxinemia and developmental abnormalities in female offspring. *Toxicol Sci*. 2017;155(2):409-419.
84. Domínguez A, Salazar Z, Arenas E, et al. Effect of perfluorooctane sulfonate on viability, maturation and gap junctional intercellular communication of porcine oocytes in vitro. *Toxicol In Vitro*. 2016;35:93-99.
85. Hallberg I, Kjellgren J, Persson S, Örn S, Sjunnesson Y. Perfluorononanoic acid (PFNA) alters lipid accumulation in bovine blastocysts after oocyte exposure during in vitro maturation. *Reprod Toxicol*. 2019;84:1-8.
86. López-Arellano P, López-Arellano K, Luna J, et al. Perfluorooctanoic acid disrupts gap junction intercellular communication and induces reactive oxygen species formation and apoptosis in mouse ovaries. *Environ Toxicol*. 2019;34(1):92-98.
87. Elcombe CR, Elcombe BM, Foster JR, et al. Hepatocellular hypertrophy and cell proliferation in Sprague-Dawley rats following dietary exposure to ammonium perfluorooctanoate occurs through increased activation of the xenosensor nuclear receptors PPAR α and CAR/PXR. *Arch Toxicol*. 2010;84(10):787-798.
88. Keller H, Givel F, Perroud M, Wahli W. Signaling cross-talk between peroxisome proliferator-activated receptor/retinoid X receptor and estrogen receptor through estrogen response elements. *Mol Endocrinol*. 1995;9(7):794-804.
89. Fan W, Yanase T, Morinaga H, et al. Activation of peroxisome proliferator-activated receptor-gamma and retinoid X receptor inhibits aromatase transcription via nuclear factor-kappaB. *Endocrinology*. 2005;146(1):85-92.
90. Rak-Mardyła A, Karpeta A. Rosiglitazone stimulates peroxisome proliferator-activated receptor gamma expression and directly affects in vitro steroidogenesis in porcine ovarian follicles. *Theriogenology*. 2014;82(1):1-9.
91. Shi Z, Zhang H, Ding L, Feng Y, Xu M, Dai J. The effect of perfluorododecanoic acid on endocrine status, sex hormones and expression of steroidogenic genes in pubertal female rats. *Reprod Toxicol*. 2009;27(3-4):352-359.
92. Wang X, Bai Y, Tang C, Cao X, Chang F, Chen L. Impact of perfluorooctane sulfonate on reproductive ability of female mice through suppression of estrogen receptor α -activated kisspeptin neurons. *Toxicol Sci*. 2018;165(2):475-486.
93. Benninghoff AD, Bisson WH, Koch DC, Ehresman DJ, Kolluri SK, Williams DE. Estrogen-like activity of perfluoroalkyl acids in vivo and interaction with human and rainbow trout estrogen receptors in vitro. *Toxicol Sci*. 2011;120(1):42-58.
94. Henry ND, Fair PA. Comparison of in vitro cytotoxicity, estrogenicity and anti-estrogenicity of triclosan, perfluorooctane sulfonate and perfluorooctanoic acid. *J Appl Toxicol*. 2013;33(4):265-272.
95. Du G, Huang H, Hu J, et al. Endocrine-related effects of perfluorooctanoic acid (PFOA) in zebrafish, H295R steroidogenesis and receptor reporter gene assays. *Chemosphere*. 2013;91(8):1099-1106.
96. Cain KC, Harlow SD, Little RJ, et al. Bias due to left truncation and left censoring in longitudinal studies of developmental and disease processes. *Am J Epidemiol*. 2011;173(9):1078-1084.